

Neural Systems Controlling the Drive to Eat: Mind Versus Metabolism

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With the bleak outlook that 75% of Americans will be overweight or obese in 10 years, it is essential to find efficient help very soon. Knowledge of the powerful and complex neural systems conferring the basic drive to eat is a prerequisite for designing efficient therapies. Recent studies suggest that the cross talk between brain areas involved in cognitive, emotional, and metabolic-regulatory functions may explain why energy homeostasis breaks down for many predisposed individuals in our modern environment.

The obesity epidemic continues unabated (47, 48, 55, 84, 87). It causes secondary health risks such as Type 2 diabetes, cardiovascular disease, sleep apnea, and depression that significantly reduce the quality of life and burden the public health system. With little or no effective treatments available, it is important to come to a better understanding of the controls of appetite and body weight and the major factors contributing to their breakdown in obesity. Genetic predisposition and physiology are important determinants of proper functioning of these control systems, but clearly the changing environment and lifestyle are the primary engines of the obesity epidemic.

Essential Physiological Functions Require Powerful and Foolproof Control Systems

Given its vital importance, procurement of sufficient energy and essential nutrients is defended by a complex system consisting of redundant pathways to make it fail-safe. The major components of this system are accurate sensors of the internal milieu and the external world, flexible and adaptive integrators that make sense out of all this diverse input, and powerful effectors that act on both the input and the output arm of energy balance (FIGURE 1). Research efforts in the post-leptin discovery era have mainly focused on the “metabolic brain,” identifying some of the crucial neural circuits in hypothalamus and hindbrain. However, to understand how metabolic need is translated into strong behavioral actions that successfully compete with other motivated behavior, the role of the “cognitive and emotional brain” cannot be neglected. Investigating how the two domains interact with each other might be particularly fruitful.

Ingested Nutrients Generate a Host of Signals Used by the Brain and Peripheral Organs

To make decisions, the brain keeps track of internal nutrient availability, a process often referred to as

nutrient sensing. Although nutrients circulating in the blood satisfy immediate needs and are sensed continuously, information from stored nutrients and nutrients ready to be absorbed from the gut are in many ways more important, because they safeguard supply over longer time periods and decrease vulnerability to external factors.

Nutrients in the gut signal future availability and satiation

Both the absence and presence of nutrients in the gastrointestinal tract generates powerful signals informing the brain and other peripheral organs (FIGURE 2). Ghrelin secreted from the mucosa of the empty stomach is the only known hormone to stimulate ingestive behavior in a feed-forward fashion (22, 23). Its plasma levels peak just before meals are requested and decline rapidly after nutrients enter the duodenum. Ghrelin may be the explanation for “hunger pangs” thought to originate from the empty stomach mucosa rubbing against each other. Ghrelin not only affects the homeostatic energy regulator in the hypothalamus but also has widespread effects on brain areas important for efficient foraging behavior (see below).

All other known gut signals essentially produce satiation of hunger, with little evidence that their absence reciprocally stimulates hunger. In the stomach, such signals include mechanical distension of vagal stretch and tension receptors and locally produced leptin acting on vagal afferents (4, 65, 86, 91). The bowel generates a number of nutrient-related signals with some degree of macronutrient-specific encoding as an emerging principle (FIGURE 2). Although fat and protein appear to signal primarily through the release of CCK and perhaps PYY(3-36) (34, 43, 68, 76), glucose signaling may involve the sweet-specific G-protein-coupled taste receptor-1 heterodimers T1R2+3, the sodium-glucose transporter, and ultimately the release of 5-HT and GLP-1 from enteroendocrine cells (31, 38, 49, 50). It will be interesting to see whether “taste receptors” are more generally used by the gut to drive nutrient-specific patterns of hormone release, thus encoding the macronutrient content of the gut. It

will be equally important to fully characterize other factors modulating hormone release from enteroendocrine cells, such as neural inputs from the enteric and autonomic nervous systems, paracrine inputs from neighboring enterocytes, and humoral inputs from the circulation. Perhaps, together with gastric volume sensors, intestinal nutrient sensors are able to quantitatively meter the gut content of each macronutrient available for absorption during the few hours after consumption.

Circulating nutrients signal immediate availability

Not only the brain, but also the pancreas, is interested in information regarding about-to-be-absorbed nutrients, since it secretes hormones important for efficient partitioning and stabilization of plasma levels of nutrients. This role of gut hormones on the endocrine pancreas was coined the incretin effect (FIGURE 2).

Together with circulating metabolites, incretins determine pancreatic secretion of a number of hormones that, in addition to their many peripheral actions, powerfully affect the brain. Given the exquisite glucose-sensitivity of the endocrine pancreatic hormone insulin and the co-secreted amylin, these two hormones provide important information regarding availability of the preferred brain fuel, glucose.

Although most of these signals act directly in the brain, some act on vagal sensory neurons innervating the portal vein and liver (12, 14, 15, 60). Particularly, a vagal sensor in the wall of the portal vein sensitive to circulating glucose and GLP-1 has gained attention since it may mediate the satiating effects of intestinal gluconeogenesis and GLP-1 (54) (FIGURE 2).

Stored nutrients signal long-term availability

White adipose tissue, far from being a passive fat storage organ, secretes a plethora of hormones and cytokines, some of which powerfully affect appetite and energy balance (80). Leptin is the most infamous hormone in energy balance regulation but has not turned out to be the magic bullet for the treatment of obesity. Although average plasma levels of leptin are positively correlated with total adiposity, fasting can rapidly suppress and re-feeding can rapidly augment leptin release, suggesting that plasma levels are rather tracking the acute nutritional state than signaling adiposity. The respective roles of the sympathetic nervous system and circulating nutrients and hormones in leptin secretion are still not completely understood.

Key Neurons in the Hypothalamus Integrate Metabolic Signals and Engage Specific Effector Pathways

Sensors and receptors for nutrition-relevant molecules are located in all major brain areas (FIGURE 2). Presence of receptors for a particular hormone typically leads to the site of its action. This is generally true for the brain too, except that some peptide hormones are also produced within the brain, making it difficult to distinguish the two sources of ligand and their specific sites of action. This is exemplified by GLP-1, which is produced not only in the lower gut but also in a population of neurons in the medulla with projections to the hypothalamus and receptors distributed in both of these brain areas.

Most molecules relevant for energy balance use specific transport systems to get across the blood-brain barrier, but there are several holes in this barrier, most prominently in the median eminence of the hypothalamus and the area postrema in the medulla oblongata (6, 7). Fully understanding the capacity and modulatory factors of these transporters may provide important tools for manipulating brain levels of relevant molecules.

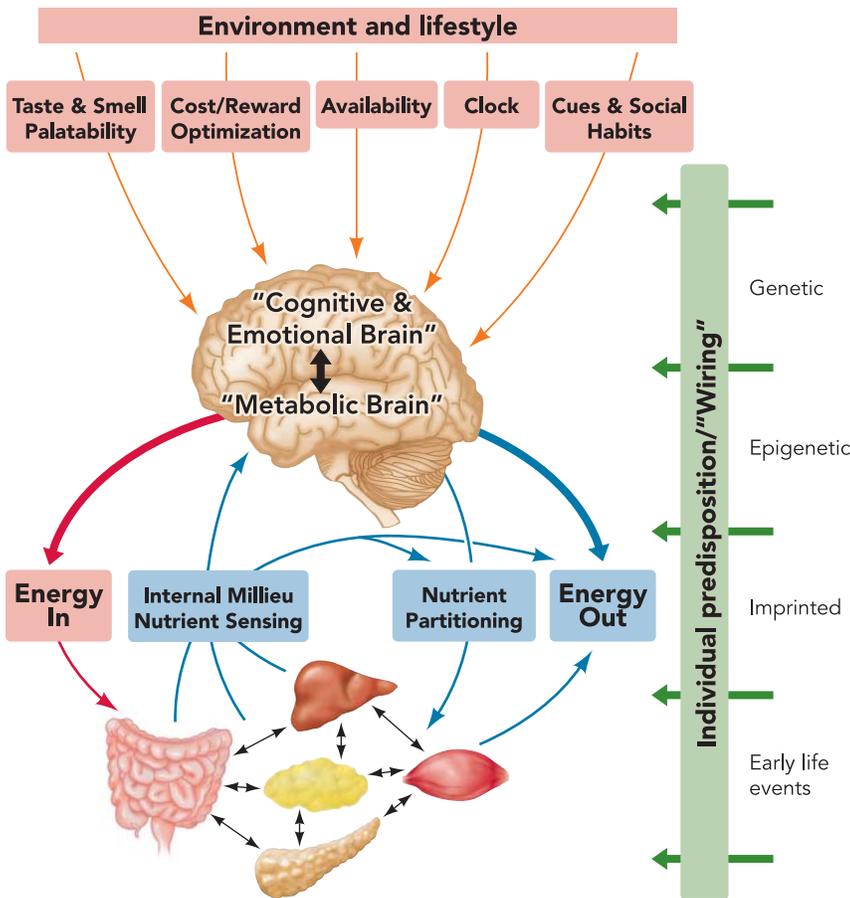


FIGURE 1. Schematic diagram showing the major factors determining neural control of appetite and regulation of energy balance

The brain monitors the internal milieu through a number of hormonal- and neural nutrient-sensing mechanisms and is under constant influence of the environment and lifestyle through the senses and mainly the cognitive and emotional brain. The two streams of information are integrated to generate adaptive behavioral (food intake) and autonomic/endocrine responses determining nutrient partitioning, energy expenditure, and overall energy balance. Any of the peripheral and central signaling steps are subject to individual predisposition through either genetic, epigenetic, or non-genetic early life imprinting mechanisms.

NPY/AGRP and POMC/CART neurons: orchestrating survival and handling abundance

The neurocircuitry involved in energy balance regulation has been discussed in several in-depth reviews (12a, 33) and is, therefore, only briefly discussed here. Two populations of neurons, one expressing the powerfully orexigenic peptides NPY and AGRP and the other expressing the anorexigenic peptides POMC and CART, located in the arcuate nucleus of the hypothalamus are the primary integrators of various nutritional information (FIGURE 3). Both populations are directly and differentially sensitive not just to circulating leptin, but also to other hormones including insulin, ghrelin, and PYY(3-36), as well as to circulating metabolites including glucose, fatty acids, and amino acids (2, 20, 62).

Since progress has been made in defining the intracellular signaling pathways mediating the effects of these various hormonal, transmitter, and metabolite signals, it becomes apparent that the richness of these intracellular signaling pathways and their coupling to changes in neuronal excitability, peptide expression, and synaptic connectivity may provide the major substrate for integrative processes (FIGURE 3). Leptin

engages a number of intracellular pathways, including those associated with cAMP, MAPK, Stat3, and PI3K (21, 56, 58, 61, 67, 73, 74, 77, 92).

AMPK, an evolutionarily conserved serine-threonine kinase activated by a high AMP-to-ATP ratio, indicating cellular energy depletion (90), stimulates food intake (53) and appears to be important for glucose sensing of POMC and AGRP neurons but not for the effects of leptin (18).

Mammalian target of rapamycin (mTOR) is another energy sensor that has assumed a specific functional role in hypothalamic regulation of energy balance. It is essential for the ability of amino acids to suppress food intake and regulate neuropeptide expression, and it may contribute to leptin and insulin action (20, 57). Since it also interacts with AMPK(51), mTOR may represent a potential site of convergence for both hormonal and nutrient sensing.

NPY/AgRP and POMC neurons interact on several levels. One is inhibition of POMC neurons via local axon collaterals from NPY neurons (21, 71). Since NPY neurons can also produce GABA, if activated, they inhibit POMC neurons through both NPY1 and GABA_A receptors. In the absence of reciprocal inhibition of NPY neurons by POMC neurons, this

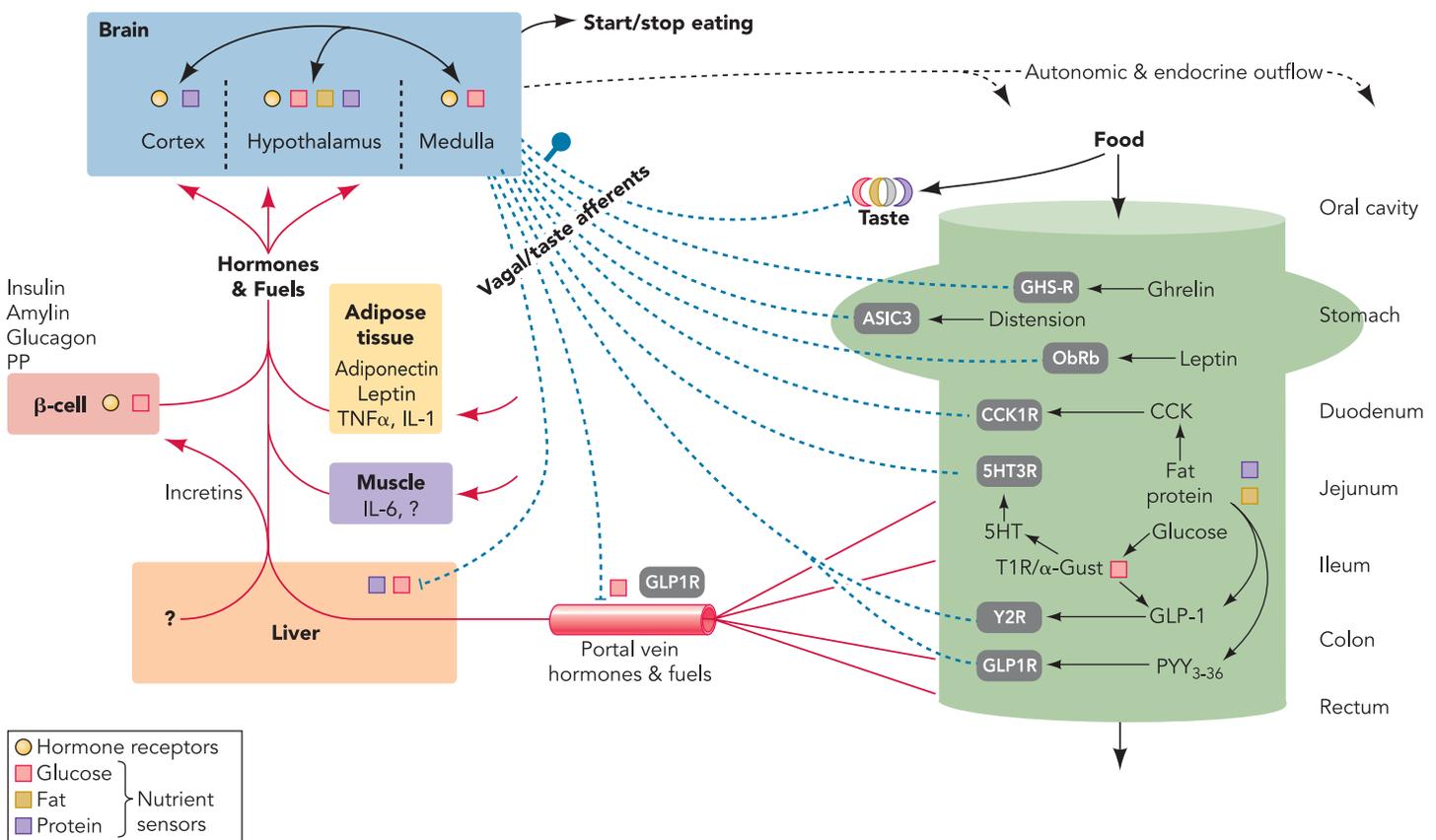


FIGURE 2. Nutrient sensing by the brain

Simplified schematic diagram showing the major pre- and postabsorptive transduction sites and mechanisms for the detection of ingested food and its macronutrient components. Nutrient information is sent to the brain through vagal and taste afferents (heavy broken lines) or through the blood circulation (solid lines). Specific receptors expressed by vagal afferent neurons are shown in rectangular boxes. Specific sensor mechanisms demonstrated for glucose, amino acids/proteins, and lipids/fatty acids are shown by gray, striped, and white squares, respectively.

arrangement may be interpreted as a fail-safe system to protect eating as the default mode of action. A second level of interaction occurs at downstream target neurons, since NPY/AgRP and POMC projections frequently converge on common downstream sites both within and outside the hypothalamus.

Autonomic, endocrine, and behavioral effector pathways

The major projection sites are second-order neuron populations in the lateral/perifornical hypothalamic area (LHA) and the paraventricular nucleus of the hypothalamus (PVH). These two brain regions are classically associated with the regulation of food intake and autonomic output, and each contains a variety of neuropeptide-expressing neurons associat-

ed with energy balance control. The prevailing model suggests that input from NPY/AgRP neurons is opposed by input from POMC neurons; this “metabolic” information is integrated with input from additional brain areas, and these downstream neurons in turn project widely to third and higher order neurons located in many areas of the brain and spinal cord (11).

Neurons within the LHA receiving direct input from the arcuate nucleus contain several food regulatory neuropeptides including orexin/hypocretin (HCRT), melanin-concentrating hormone (MCH), cocaine and amphetamine-regulated transcript (CART), neurtensin (NT), and histamine (FIGURE 3). Specific populations of LHA neurons also express leptin receptors, and some are sensitive to glucose. In addition to this metabolic information, the LHA also receives

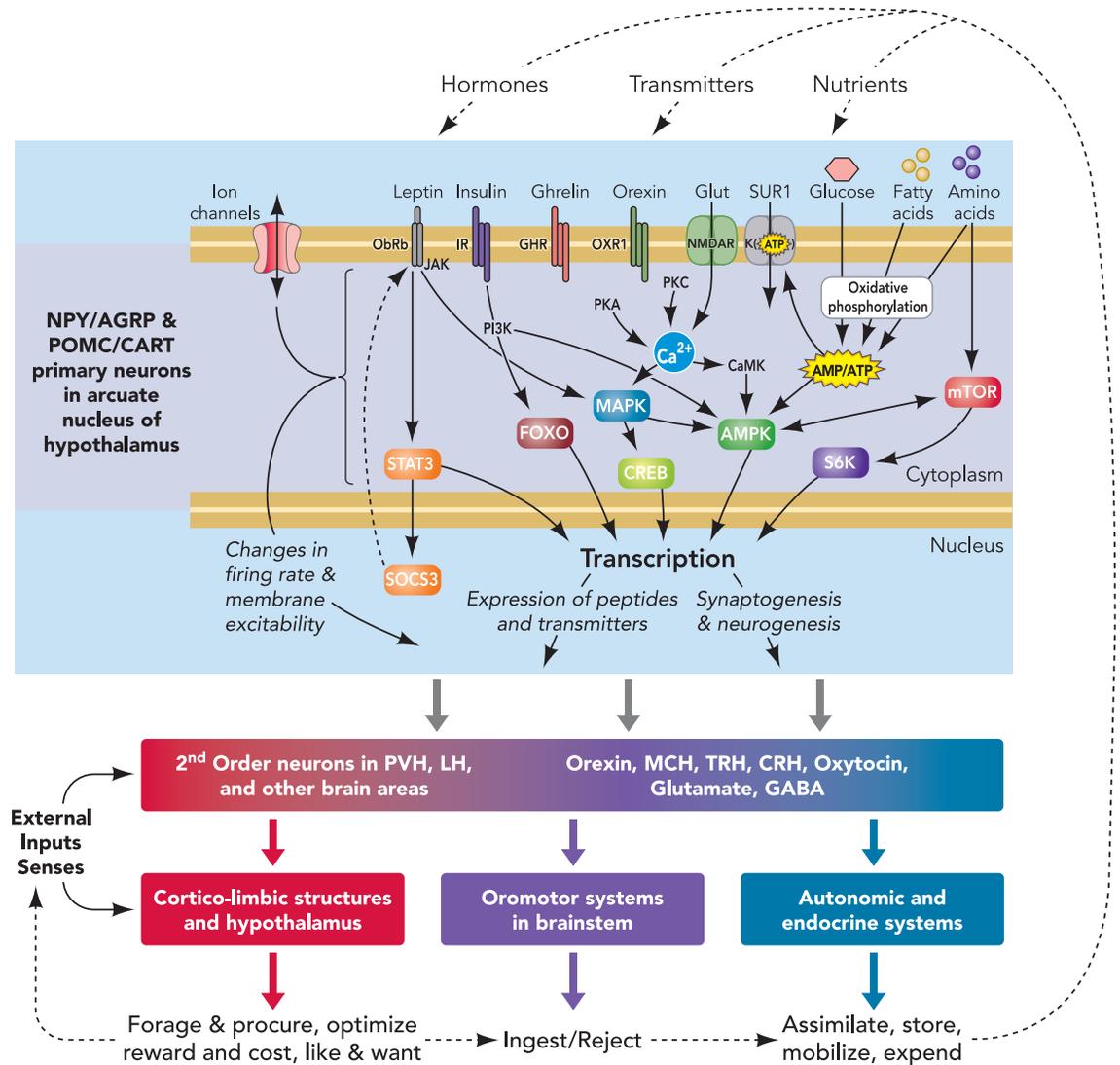


FIGURE 3. Hypothalamic regulator of energy balance

Highly schematic diagram depicting the major purported intracellular signaling pathways in a hypothetical arcuate nucleus neuron that integrates nutrient, hormonal, and neurotransmitter signals and activates downstream neural networks leading to behavioral, autonomic, and endocrine responses. ObRb, long form of leptin receptor; IR, insulin receptor; GHR, ghrelin receptor; OXR1, orexin-type-1 receptor; Glut, glutamate; NMDAR, NMDR-glutamate receptor; SUR1, sulfonylurea receptor; JAK, janus kinase; AMPK, adenosine monophosphate-dependent kinase; mTOR, mammalian target of rapamicin; PKA, protein kinase A.

information from brain areas associated with reward, motivation, learning, and memory and from brain stem areas associated with vagal and visceral sensory input, sensorimotor coordination, and arousal. In turn, these peptidergic second-order LHA neurons project widely through the entire brain (11), from the cortex to the spinal cord (FIGURE 4).

Second-order neurons in the PVH are classically associated with autonomic and neuroendocrine functions. Thyrotropin-releasing hormone (TRH) and corticotropin-releasing hormone (CRH) neurons receive direct input from both types of ARC neurons and regulate the thyroid and HPA axis and stress response, respectively (45).

In addition to the LHA and PVH, arcuate POMC/CART neurons also project to other brain areas. For example, leptin-sensitive POMC neurons project directly to brain stem areas associated with the response to satiety signals and autonomic outflow (13, 93).

What is homeostatic regulation?

The hypothalamic neurocircuitry discussed above is crucial to energy homeostasis as indicated by the development of obesity or leanness after loss- or gain-of-function manipulations of its main components.

Although this circuit is assumed by many to regulate body weight and adiposity within a narrow set point, much like a thermostat controls room temperature, this view has been largely abandoned in favor of a more flexible regulator that can learn from past experience and adapt to changing environmental factors. Arguably, the major force “designing” the system was the constant struggle throughout evolution to find enough food for survival, resulting in a very strong defense of the lower limits of adiposity. One school of thought is that, over the millions of years of evolution, genes optimizing foraging and fuel efficiency were selected at least in some populations (66). Clearly, evolutionary pressure has also existed to limit the upper limits of adiposity and, perhaps more likely, body weight (78). Disadvantages of elevated body weight are particularly evident in birds and in becoming prey because of slower running speed.

Cortico-Limbic Pathways Coordinate Metabolic Need with the External World

It is clear that the neurocircuitry originating from the primary energy sensors in the arcuate nucleus

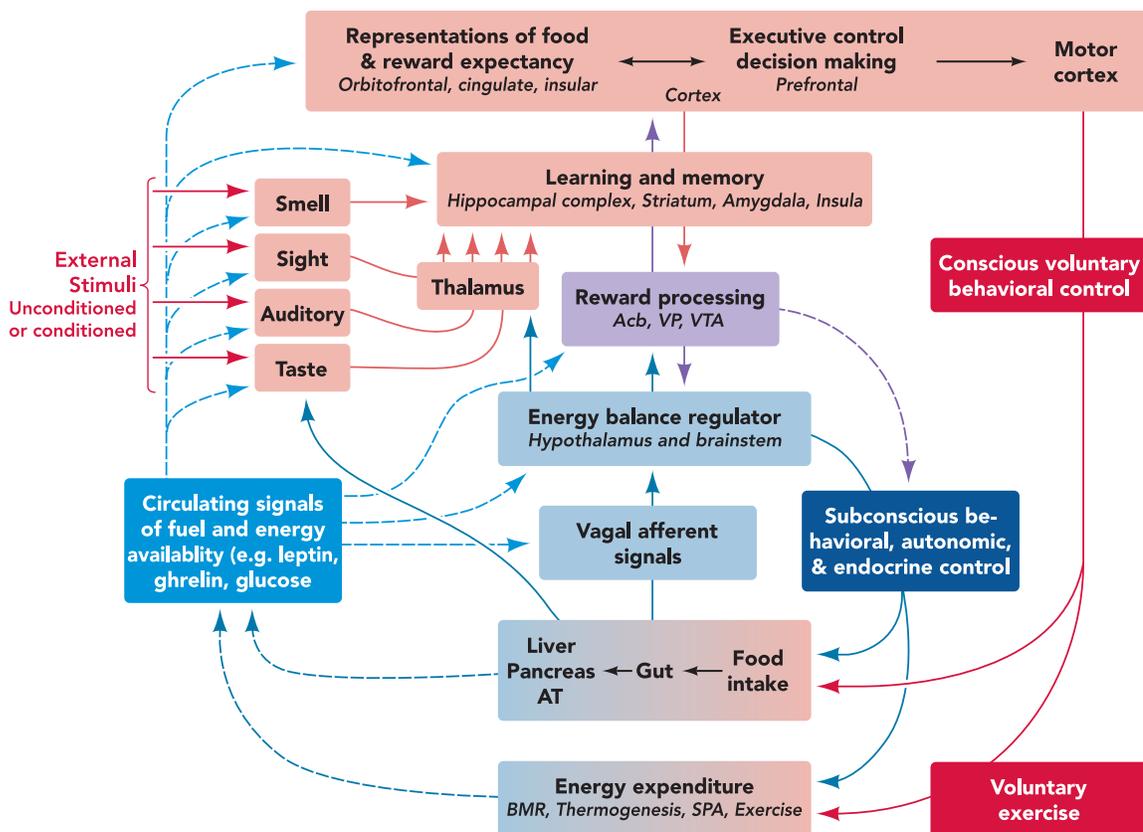


FIGURE 4. Major systems and pathways responsible for the neural integration of internal and external information in the control of appetite and energy expenditure
 Blue areas and pathways are mainly involved in metabolic and energy balance regulation. Red areas and pathways are mainly involved in communication with the external world through cognitive and emotional processes such as learning and memory, reward, mood, stress, choice, and decision making.

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described above is embedded in a much larger neural system that allows adaptation and coordination of metabolic needs to the demands and intricacies of the prevailing environment. For example, it does not make sense for the hungry vole to leave the burrow if the weasel waits outside. In fact, much of the brain has evolved to take care of hunger ever since mobile life forms emerged. Although procuring food in our modern environment is as easy as grabbing a piece of birthday cake on the table, it used to be a demanding task for most of the last 5 million years. Even more primitive invertebrate animals such as honey bees and ants use elaborate navigation and communication strategies to secure food sources and guarantee survival (30, 70, 85).

“Although humans have the ability to make conscious, voluntary decisions and choices, most of our actions have a subconscious component that escapes voluntary control.”

Representations of experience with food

We remember past experiences with foods, particularly if the experience was out of the ordinary. Experiences that evoked either extreme pleasure or complete disgust generate the most salient memories. Thus we remember the restaurant and everything in and around it very well, where we had that extraordinary dish, and we remember even an average dish when we fell in love at that occasion. On the other hand, we immediately recognize and avoid a food that made us sick. A growing number of studies suggest that representations of experience with foods are generated in the orbitofrontal cortex, an area in the prefrontal cortex that receives converging information through all sensory modalities (FIGURE 4) (83). Therefore, representations contain a number of sensory attributes, including shape, color, taste, and flavor, as well as links to time, location, social context, cost, and reward expectation (25, 83).

It is not clear how and where exactly such representations are stored. The orbitofrontal cortex is in intimate contact with other cortical areas, particularly the anterior cingulate, perirhinal, and entorhinal cortices, as well as with the hippocampal formation and the amygdala, often collectively referred to as paralimbic cortex (for review, see Ref. 83). It is within these areas that polymodal representations are thought to be available as working memory for constant updating.

Reward and emotions

It is thought that emotions evolved as a mechanism to reinforce beneficial and suppress potentially harmful

stimuli and behaviors. For example, the sweet taste of certain foods is associated with positive emotions that augment the motivational drive to obtain such foods; in brief, it is said to be rewarding. The reward value of a particular food is bundled with the other attributes into the stored representations discussed above. Thus life is all about learning how specific behavioral responses or actions lead to positive emotions or reward in the future. However, reward is a fuzzy psychological construct and is neurologically ill-defined. Berridge has proposed parsing reward into liking, wanting, and learning, each representing separate but interlinked psychological processes with distinguishable underlying neurological substrates (10).

Liking is also known as hedonic value of a food stimulus. Its most primitive form of expression is the characteristic “happy face” expressed by rodents, primates, and humans when tasting sweets. Current knowledge suggests that liking is neurologically organized by a widely distributed system with the mu-opioid and perhaps the cannabinoid receptor systems playing common denominators (19). Since the “happy face” is observed in decerebrate rats and anencephalic human infants, neural circuits in the hindbrain appear to be sufficient for the basic expression of liking (36, 79). In addition, areas in the ventral striatum and the amygdala are undoubtedly part of this distributed neural network of liking. To consciously experience and give subjective ratings of pleasure from palatable foods (liking), humans appear to also use areas in the prefrontal and cingulate cortex (42).

Wanting, or incentive salience, is another component of reward as proposed by Berridge (9, 10). It usually, but not always, follows liking. Although liking is closer to sensory processes, wanting is closer to motor action. Wanting can be further subdivided into decision-making and motor action. It can be shown that activity of the mesolimbic dopamine system accurately reflects the cue-driven inclination to choose one behavior over another to optimize reward. Neurologically, wanting is intimately linked to the mesolimbic dopamine system, which is crucial for the orchestration of motor behavior to obtain rewards. Dopaminergic projections from the ventral tegmental area to the nucleus accumbens and prefrontal cortex are the most important component of the implicit or unconscious “wanting” system (FIGURE 4) (24, 41, 89). Manipulation of this dopamine system powerfully influences wanting (instrumental performance for and consumption of) drugs or food but not “liking” (10, 16, 63, 88).

Metabolic signals modulate all levels of food-related cognitive and reward processing

It was originally thought that the classic nutritional feedback signals, such as leptin, insulin, gut hormones, and circulating nutrients themselves, act mainly on a few areas of the brain such as specific

parts of the hypothalamus and brain stem (see above). However, recent studies suggest that these metabolic signals have a much broader influence on brain functions (FIGURE 4).

For example, leptin has been shown to modulate food-related sensory input signals of all modalities, even at early stages of processing, so that low leptin levels can dramatically lower detection thresholds of external stimuli signaling availability of nutrients (35, 40, 75, 82). Leptin and insulin can also act directly on mesolimbic dopamine neurons to modulate wanting of food (29, 32, 37). Neural activity in the nucleus accumbens elicited by visual food stimuli is very high in genetically leptin-deficient adolescents and promptly returns to normal levels on leptin administration. While in the leptin deficient state, nucleus accumbens activation was positively correlated with ratings of liking in both the fasted and fed state; it was correlated only in the fasted state after leptin treatment and in normal individuals (28). The lower gut hormone PYY(3-36), which has now been convincingly demonstrated to suppress food intake in humans and rodents (17), also modulates activity of the ventral tegmental area (VTA) and ventral striatum (8). In contrast to leptin, the gut hormone ghrelin appears to facilitate foraging behavior and increase reward processing as part of its orexigenic action (1, 27, 39, 72).

Conscious decisions vs. subconscious drive

Although humans have the ability to make conscious, voluntary decisions and choices, most of our actions have a subconscious component that escapes voluntary control. This is why we eat palatable food such as chocolate in the absence of any metabolic need, even if we “know” better not to do it. The right prefrontal cortex appears to play a critical role in behavioral restraint and moral self-control by keeping reward-generating mechanisms in check (3, 46). The prefrontal cortex receives sensory information from inside and outside the body as well as emotional and cognitive information from the limbic system, and it is intimately connected to cortical areas involved in motor planning and execution. It is thus in an ideal position to translate all available homeostatic and environmental information into adaptive behavioral responses, in brief, to make choices and decisions (5, 59). Damage to the right frontal cortex can lead to a general disregard for the long-term adverse consequences of behavioral choices, such as increased risk taking and excessive food intake (3). A “Gourmand syndrome” with passion for eating highly palatable foods was reported in two case studies of humans with damage to the right frontal hemisphere (69, 81).

Modern neuroimaging studies also support the importance of a balanced control by distinct areas of the prefrontal cortex in the control of food intake.

Successful dieters who have significantly higher levels of dietary restraint to nondieters show increased neural activity in the right dorsolateral prefrontal cortex in response to food consumption (26). In contrast, obese subjects show less activation of the left dorsolateral prefrontal cortex in response to food (44), and patients suffering from the Prader-Willi syndrome, who show severe disturbances in appetite control resulting in hyperphagia and obesity, show increased activity in the ventromedial prefrontal cortex when viewing pictures of food after glucose consumption (52). This latter finding is consistent with a role of the ventromedial prefrontal cortex in the mediation of food intake driven by conditioned (learned) motivational cues in sated rats (64).

Conclusions and Perspectives

The neural systems conferring the basic drive to eat are powerful, distributed, and redundant, and have been relatively refractory to pharmacological intervention and prevention of obesity. This is because in most obese people there is not one identifiable single defect in this system that could be corrected by drugs. Rather, there is a mismatch between the “thrifty” evolution of a large number of genes and their products and the current environment of plenty and sedentary lifestyle. Thus it is unlikely that mono-therapies, only acting on one single step in one pathway, will be successful in fighting the obesity war. Combination therapies acting on several steps in several pathways and involving lifestyle changes look more promising. Currently, the most effective treatment for morbid obesity is surgery, and it appears that Roux-en Y gastric bypass surgery “works” by favorably changing the physiological cross talk between the gut and the brain rather than through mechanical hindrance and malabsorption. So, unexpectedly, this procedure may help us understand what the critical pathways are that need to be simultaneously manipulated to trick the wisdom of the body.

References

1. Abizaid A, Liu ZW, Andrews ZB, Shanabrough M, Borok E, Elsworth JD, Roth RH, Sleeman MW, Picciotto MR, Tschop MH, Gao XB, Horvath TL. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *J Clin Invest* 116: 3229–3239, 2006.
2. Akabayashi A, Zaia CT, Silva I, Chae HJ, Leibowitz SF. Neuropeptide Y in the arcuate nucleus is modulated by alterations in glucose utilization. *Brain Res* 621: 343–348, 1993.
3. Alonso-Alonso M, Pascual-Leone A. The right brain hypothesis for obesity. *JAMA* 297: 1819–1822, 2007.
4. Bado A, Levasseur S, Attoub S, Kermorgant S, Laigneau JP, Bortoluzzi MN, Moizo L, Lehy T, Guerre-Millo M, Le Marchand-Brustel Y, and Lewin MJ. The stomach is a source of leptin. *Nature* 394: 790–793, 1998.
5. Balleine BW. The neural basis of choice and decision making. *J Neurosci* 27: 8159–8160, 2007.
6. Banks WA. Is obesity a disease of the blood-brain barrier? Physiological, pathological, and evolutionary considerations. *Curr Pharm Des* 9: 801–809, 2003.

7. Banks WA, Farr SA, Morley JE. The effects of high fat diets on the blood-brain barrier transport of leptin: failure or adaptation? *Physiol Behav* 88: 244–248, 2006.
8. Batterham RL, fytche DH, Rosenthal JM, Zelaya FO, Barker GJ, Withers DJ, Williams SC. PYY modulation of cortical and hypothalamic brain areas predicts feeding behaviour in humans. *Nature* 450: 106–109, 2007.
9. Berridge KC. Food reward: brain substrates of wanting and liking. *Neurosci Biobehav Rev* 20: 1–25, 1996.
10. Berridge KC, Robinson TE. Parsing reward. *Trends Neurosci* 26: 507–513, 2003.
11. Berthoud HR. Multiple neural systems controlling food intake and body weight. *Neurosci Biobehav Rev* 26: 393–428, 2002.
12. Berthoud HR, Kressel M, Neuhuber WL. An anterograde tracing study of the vagal innervation of rat liver, portal vein and biliary system. *Anat Embryol (Berl)* 186: 431–442, 1992.
- 12a. Bethoud HR, Morrison C. The brain, appetite, and obesity. *Annu Rev Psychol* 59: 55–92, 2008.
13. Berthoud HR, Sutton GM, Townsend RL, Patterson LM, Zheng H. Brainstem mechanisms integrating gut-derived satiety signals and descending forebrain information in the control of meal size. *Physiol Behav* 89: 517–524, 2006.
14. Burcelin R, Da Costa A, Drucker D, Thorens B. Glucose competence of the hepatoportal vein sensor requires the presence of an activated glucagon-like peptide-1 receptor. *Diabetes* 50: 1720–1728, 2001.
15. Burcelin R, Dolci W, Thorens B. Glucose sensing by the hepatoportal sensor is GLUT2-dependent: in vivo analysis in GLUT2-null mice. *Diabetes* 49: 1643–1648, 2000.
16. Cannon CM, Palmiter RD. Reward without dopamine. *J Neurosci* 23: 10827–10831, 2003.
17. Chelikani PK, Haver AC, Reeve JR Jr, Keire DA, and Reidelberger RD. Daily, intermittent intravenous infusion of peptide YY(3-36) reduces daily food intake and adiposity in rats. *Am J Physiol Regul Integr Comp Physiol* 290: R298–R305, 2006.
18. Claret M, Smith MA, Batterham RL, Selman C, Choudhury AI, Fryer LG, Clements M, Al-Qassab H, Heffron H, Xu AW, Speakman JR, Barsh GS, Viollet B, Vaulont S, Ashford ML, Carling D, Withers DJ. AMPK is essential for energy homeostasis regulation and glucose sensing by POMC and AgRP neurons. *J Clin Invest* 117: 2325–2336, 2007.
19. Cooper SJ. Endocannabinoids and food consumption: comparisons with benzodiazepine and opioid palatability-dependent appetite. *Eur J Pharmacol* 500: 37–49, 2004.
20. Cota D, Proulx K, Smith KA, Kozma SC, Thomas G, Woods SC, Seeley RJ. Hypothalamic mTOR signaling regulates food intake. *Science* 312: 927–930, 2006.
21. Cowley MA, Smart JL, Rubinstein M, Cerdan MG, Diano S, Horvath TL, Cone RD, Low MJ. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 411: 480–484, 2001.
22. Cummings DE. Ghrelin and the short- and long-term regulation of appetite and body weight. *Physiol Behav* 89: 71–84, 2006.
23. Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 50: 1714–1719, 2001.
24. Dayan P, Balleine BW. Reward, motivation, and reinforcement learning. *Neuron* 36: 285–298, 2002.
25. de Araujo IE, Rolls ET, Velazco MI, Margot C, Cayeux I. Cognitive modulation of olfactory processing. *Neuron* 46: 671–679, 2005.
26. DelParigi A, Chen K, Salbe AD, Hill JO, Wing RR, Reiman EM, Tataranni PA. Successful dieters have increased neural activity in cortical areas involved in the control of behavior. *Int J Obes* 31: 440–448, 2007.
27. Diano S, Farr SA, Benoit SC, McNay EC, da Silva I, Horvath B, Gaskin FS, Nonaka N, Jaeger LB, Banks WA, Morley JE, Pinto S, Sherwin RS, Xu L, Yamada KA, Sleeman MW, Tschop MH, Horvath TL. Ghrelin controls hippocampal spine synapse density and memory performance. *Nat Neurosci* 9: 381–388, 2006.
28. Farooqi IS, Bullmore E, Keogh J, Gillard J, O'Rahilly S, Fletcher PC. Leptin regulates striatal regions and human eating behavior. *Science* 317: 1355, 2007.
29. Figlewicz DP. Adiposity signals and food reward: expanding the CNS roles of insulin and leptin. *Am J Physiol Regul Integr Comp Physiol* 284: R882–R892, 2003.
30. Franks NR, Richardson T. Teaching in tandem-running ants. *Nature* 439: 153, 2006.
31. Freeman SL, Bohan D, Darcel N, Raybould HE. Luminal glucose sensing in the rat intestine has characteristics of a sodium-glucose cotransporter. *Am J Physiol Gastrointest Liver Physiol* 291: G439–G445, 2006.
32. Fulton S, Pissios P, Manchon RP, Stiles L, Frank L, Pothos EN, Maratos-Flier E, Flier JS. Leptin regulation of the mesoaccumbens dopamine pathway. *Neuron* 51: 811–822, 2006.
33. Gao Q, Horvath TL. Neurobiology of feeding and energy expenditure. *Annu Rev Neurosci* 30: 367–398, 2007.
34. Geary N. Endocrine controls of eating: CCK, leptin, and ghrelin. *Physiol Behav* 81: 719–733, 2004.
35. Getchell TV, Kwong K, Saunders CP, Stromberg AJ, Getchell ML. Leptin regulates olfactory-mediated behavior in ob/ob mice. *Physiol Behav* 87: 848–856, 2006.
36. Grill HJ, Norgren R. The taste reactivity test. II. Mimetic responses to gustatory stimuli in chronic thalamic and chronic decerebrate rats. *Brain Res* 143: 281–297, 1978.
37. Hommel JD, Trinko R, Sears RM, Georgescu D, Liu ZW, Gao XB, Thurmon JJ, Marinelli M, DiLeone RJ. Leptin receptor signaling in midbrain dopamine neurons regulates feeding. *Neuron* 51: 801–810, 2006.
38. Jang HJ, Kokrashvili Z, Theodorakis MJ, Carlson OD, Kim BJ, Zhou J, Kim HH, Xu X, Chan SL, Juhaszova M, Bernier M, Mosinger B, Margolskee RF, Egan JM. Gut-expressed gustducin and taste receptors regulate secretion of glucagon-like peptide-1. *Proc Natl Acad Sci USA* 104: 15069–15074, 2007.
39. Jerlhag E, Egecioglu E, Dickson SL, Douhan A, Svensson L, Engel JA. Ghrelin administration into tegmental areas stimulates locomotor activity and increases extracellular concentration of dopamine in the nucleus accumbens. *Addict Biol* 12: 6–16, 2007.
40. Julliard AK, Chaput MA, Apfelbaum A, Aime P, Mahfouz M, Duchamp-Viret P. Changes in rat olfactory detection performance induced by orexin and leptin mimicking fasting and satiation. *Behav Brain Res* 183: 123–129, 2007.
41. Kaczmarek HJ, Kiefer SW. Microinjections of dopaminergic agents in the nucleus accumbens affect ethanol consumption but not palatability. *Pharmacol Biochem Behav* 66: 307–312, 2000.
42. Kringelbach ML. Food for thought: hedonic experience beyond homeostasis in the human brain. *Neuroscience* 126: 807–819, 2004.
43. Lal S, Kirkup AJ, Brunnsden AM, Thompson DG, Grundy D. Vagal afferent responses to fatty acids of different chain length in the rat. *Am J Physiol Gastrointest Liver Physiol* 281: G907–G915, 2001.
44. Le DS, Pannacciulli N, Chen K, Del Parigi A, Salbe AD, Reiman EM, Krakoff J. Less activation of the left dorsolateral prefrontal cortex in response to a meal: a feature of obesity. *Am J Clin Nutr* 84: 725–731, 2006.
45. Lechan RM, Fekete C. The TRH neuron: a hypothalamic integrator of energy metabolism. *Prog Brain Res* 153: 209–235, 2006.
46. Lee D, Rushworth MF, Walton ME, Watanabe M, Sakagami M. Functional specialization of the primate frontal cortex during decision making. *J Neurosci* 27: 8170–8173, 2007.
47. Li C, Ford ES, McGuire LC, Mokdad AH. Increasing trends in waist circumference and abdominal obesity among US adults. *Obesity (Silver Spring)* 15: 216–224, 2007.
48. Li C, Ford ES, Mokdad AH, Cook S. Recent trends in waist circumference and waist-height ratio among US children and adolescents. *Pediatrics* 118: e1390–e1398, 2006.
49. Mace OJ, Affleck J, Patel N, Kellett GL. Sweet taste receptors in rat small intestine stimulate glucose absorption through apical GLUT2. *J Physiol* 582: 379–392, 2007.
50. Margolskee RF, Dyer J, Kokrashvili Z, Salmon KS, Ilegems E, Daly K, Mailliet EL, Ninomiya Y, Mosinger B, Shirazi-Beechey SP. T1R3 and gustducin in gut sense sugars to regulate expression of Na⁺-glucose cotransporter 1. *Proc Natl Acad Sci USA* 104: 15075–15080, 2007.
51. Marshall S. Role of insulin, adipocyte hormones, and nutrient-sensing pathways in regulating fuel metabolism and energy homeostasis: a nutritional perspective of diabetes, obesity, and cancer. *Sci STKE* 2006: re7, 2006.
52. Miller JL, James GA, Goldstone AP, Couch JA, He G, Driscoll DJ, Liu Y. Enhanced activation of reward mediating prefrontal regions in response to food stimuli in Prader-Willi syndrome. *J Neurol Neurosurg Psychiatry* 78: 615–619, 2007.
53. Minokoshi Y, Alquier T, Furukawa N, Kim YB, Lee A, Xue B, Mu J, Foulfelle F, Ferrer P, Birnbaum MJ, Stuck BJ, Kahn BB. AMP-kinase regulates food intake by responding to hormonal and nutrient signals in the hypothalamus. *Nature* 428: 569–574, 2004.
54. Mithieux G, Misery P, Magnan C, Pillot B, Gautier-Stein A, Bernard C, Rajas F, Zitoun C. Portal sensing of intestinal gluconeogenesis is a mechanistic link in the diminution of food intake induced by diet protein. *Cell Metab* 2: 321–329, 2005.
55. Mokdad AH, Ford ES, Bowman BA, Dietz WH, Vinicor F, Bales VS, Marks JS. Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA* 289: 76–79, 2003.
56. Morrison CD, Morton GJ, Niswender KD, Gelling RW, Schwartz MW. Leptin inhibits hypothalamic Npy and AgRP gene expression via a mechanism that requires phosphatidylinositol 3-OH-kinase signaling. *Am J Physiol Endocrinol Metab* 289: E1051–E1057, 2005.
57. Morrison CD, Xi X, White CL, Ye J, Martin RJ. Amino acids inhibit AgRP gene expression via an mTOR-dependent mechanism. *Am J Physiol Endocrinol Metab* 293: E165–E171, 2007.
58. Munzberg H, Myers MG Jr. Molecular and anatomical determinants of central leptin resistance. *Nat Neurosci* 8: 566–570, 2005.
59. Murray EA, O'Doherty JP, Schoenbaum G. What we know and do not know about the functions of the orbitofrontal cortex after 20 years of cross-species studies. *J Neurosci* 27: 8166–8169, 2007.

60. Nijima A. Glucose-sensitive afferent nerve fibres in the hepatic branch of the vagus nerve in the guinea-pig. *J Physiol* 332: 315–323, 1982.
61. Niswender KD, Morton GJ, Stearns WH, Rhodes CJ, Myers MG Jr, Schwartz MW. Intracellular signaling key enzyme in leptin-induced anorexia. *Nature* 413: 794–795, 2001.
62. Obici S, Rossetti L. Nutrient sensing and the regulation of insulin action and energy balance. *Endocrinology* 144: 5172–5178, 2003.
63. Pecina S, Cagniard B, Berridge KC, Aldridge JW, Zhuang X. Hyperdopaminergic mutant mice have higher “wanting” but not “liking” for sweet rewards. *J Neurosci* 23: 9395–9402, 2003.
64. Petrovich GD, Ross CA, Holland PC, Gallagher M. Medial prefrontal cortex is necessary for an appetitive contextual conditioned stimulus to promote eating in sated rats. *J Neurosci* 27: 6436–6441, 2007.
65. Phillips RJ, Powley TL. Tension and stretch receptors in gastrointestinal smooth muscle: re-evaluating vagal mechanoreceptor electrophysiology. *Brain Res Brain Res Rev* 34: 1–26, 2000.
66. Prentice AM, Rayco-Solon P, Moore SE. Insights from the developing world: thrifty genotypes and thrifty phenotypes. *Proc Nutr Soc* 64: 153–161, 2005.
67. Rahmouni K, Haynes WG, Morgan DA, Mark AL. Intracellular mechanisms involved in leptin regulation of sympathetic outflow. *Hypertension* 41: 763–767, 2003.
68. Raybould HE, Glatzle J, Freeman SL, Whited K, Darcel N, Liou A, Bohan D. Detection of macronutrients in the intestinal wall. *Auton Neurosci* 125: 28–33, 2006.
69. Regard M, Landis T. “Gourmand syndrome”: eating passion associated with right anterior lesions. *Neurology* 48: 1185–1190, 1997.
70. Riley JR, Greggers U, Smith AD, Reynolds DR, Menzel R. The flight paths of honeybees recruited by the waggle dance. *Nature* 435: 205–207, 2005.
71. Roseberry AG, Liu H, Jackson AC, Cai X, Friedman JM. Neuropeptide Y-mediated inhibition of pro-opiomelanocortin neurons in the arcuate nucleus shows enhanced desensitization in ob/ob mice. *Neuron* 41: 711–722, 2004.
72. Schmid DA, Held K, Ising M, Uhr M, Weikel JC, Steiger A. Ghrelin stimulates appetite, imagination of food, GH, ACTH, and cortisol, but does not affect leptin in normal controls. *Neuropsychopharmacology* 30: 1187–1192, 2005.
73. Schwartz MW, Seeley RJ, Campfield LA, Burn P, Baskin DG. Identification of targets of leptin action in rat hypothalamus. *J Clin Invest* 98: 1101–1106, 1996.
74. Schwartz MW, Seeley RJ, Woods SC, Weigle DS, Campfield LA, Burn P, Baskin DG. Leptin increases hypothalamic pro-opiomelanocortin mRNA expression in the rostral arcuate nucleus. *Diabetes* 46: 2119–2123, 1997.
75. Shigemura N, Ohta R, Kusakabe Y, Miura H, Hino A, Koyano K, Nakashima K, Ninomiya Y. Leptin modulates behavioral responses to sweet substances by influencing peripheral taste structures. *Endocrinology* 145: 839–847, 2004.
76. Smith GP, Jerome C, Norgren R. Afferent axons in abdominal vagus mediate satiety effect of cholecystokinin in rats. *Am J Physiol Regul Integr Comp Physiol* 249: R638–R641, 1985.
77. Spanswick D, Smith MA, Groppi VE, Logan SD, Ashford ML. Leptin inhibits hypothalamic neurons by activation of ATP-sensitive potassium channels. *Nature* 390: 521–525, 1997.
78. Speakman JR. Thrifty genes for obesity and the metabolic syndrome—time to call off the search? *Diab Vasc Dis Res* 3: 7–11, 2006.
79. Steiner JE. *The Gustofacial Response: Observations on Normal and Anencephalic Newborn Infants*. Bethesda, MD: US Dept. of Health, Education, and Welfare, 1973.
80. Trayhurn P, Bing C, Wood IS. Adipose tissue and adipokines: energy regulation from the human perspective. *J Nutr* 136: 1935S–1939S, 2006.
81. Uher R, Treasure J. Brain lesions and eating disorders. *J Neurol Neurosurg Psychiatry* 76: 852–857, 2005.
82. Uher R, Treasure J, Heining M, Brammer MJ, Campbell IC. Cerebral processing of food-related stimuli: effects of fasting and gender. *Behav Brain Res* 169: 111–119, 2006.
83. Verhagen JV. The neurocognitive bases of human multimodal food perception: consciousness. *Brain Res* 53: 271–286, 2006.
84. Vivian EM. Type 2 diabetes in children and adolescents: the next epidemic? *Curr Med Res Opin* 22: 297–306, 2006.
85. von Frisch K. *Dance Language and Orientation of Bees*. Cambridge, MA: Harvard Univ. Press, 1967.
86. Wang YH, Tache Y, Sheibel AB, Go VL, Wei JY. Two types of leptin-responsive gastric vagal afferent terminals: an in vitro single-unit study in rats. *Am J Physiol Regul Integr Comp Physiol* 273: R833–R837, 1997.
87. Wyatt SB, Winters KP, Dubbert PM. Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. *Am J Med Sci* 331: 166–174, 2006.
88. Wyvell CL, Berridge KC. Incentive sensitization by previous amphetamine exposure: increased cue-triggered “wanting” for sucrose reward. *J Neurosci* 21: 7831–7840, 2001.
89. Wyvell CL, Berridge KC. Intra-accumbens amphetamine increases the conditioned incentive salience of sucrose reward: enhancement of reward “wanting” without enhanced “liking” or response reinforcement. *J Neurosci* 20: 8122–8130, 2000.
90. Xue B, Kahn BB. AMPK integrates nutrient and hormonal signals to regulate food intake and energy balance through effects in the hypothalamus and peripheral tissues. *J Physiol* 574: 73–83, 2006.
91. Zagorodnyuk VP, Chen BN, Brookes SJ. Intraganglionic laminar endings are mechanotransduction sites of vagal tension receptors in the guinea-pig stomach. *J Physiol* 534: 255–268, 2001.
92. Zhao AZ, Huan JN, Gupta S, Pal R, Sahu A. A phosphatidylinositol 3-kinase phosphodiesterase 3B-cyclic AMP pathway in hypothalamic action of leptin on feeding. *Nat Neurosci* 5: 727–728, 2002.
93. Zheng H, Patterson LM, Phifer CB, Berthoud HR. Brain stem melanocortinergic modulation of meal size and identification of hypothalamic POMC projections. *Am J Physiol Regul Integr Comp Physiol* 289: R247–R258, 2005.